Studying the Elimination Pattern of Caprine Arthritis Encephalitis Virus in the Milk of Infected Females

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ABSTRACT

Background: Small ruminants can be infected by lentiviruses, such as Maedi-Visna Virus (MVV) and Caprine Arthritis-Encephalitis Virus (CAEV). The main route of transmission is via ingestion of contaminated colostrum and milk although vertical transmission can occur. Recently, several studies for molecular detection of CAEV in milk, using conventional PCR and real-time PCR are being carried out. Considering the elimination of CAEV through the milk of infected animals and the importance of this virus in the goat production, the aim of this study was to evaluate the elimination pattern of CAEV in milk, evaluating the frequency and the concentration eliminated during the lactation.

Materials, Methods & Results: A cohort of four negative females for CAEV was inseminated with semen experimentally infected with CAEV-Cork strain. They were located in stalls at the Hospital of Ruminants from School of Veterinary Medicine and Animal Science from University of São Paulo, Brazil. Goats received coast-cross hay, pellet feeding, mineral salt and water ad libitum. All females were observed every day during pregnancy. After lambing, kids received warm bovine colostrum and bovine milk powder during two months. Forty milk samples were collected at five-day interval during two months. A mixture of five milliliters from each teat was obtained and cDNA extraction was performed using DNA Mini Kit. Initially, real-time PCR was performed using an endogenous control for research of the constitutive gene (12S) for goats. Using positive samples in the first reaction, another reaction was performed using specific primers for lentiviruses based on the gag gene (conserved in retroviruses). In order to compare the results, nested-PCR was performed. After real-time PCR, cDNA was detected in samples from one female, corresponding to the day of calving, 14th, 20th, 25th, 35th and 40th day postpartum (15%; 6/40). The absence of amplified cDNA in thirty days postpartum, as well as in the final twenty days of lactation, was observed. Sample corresponding to the 7th day postpartum was not obtained. The virus concentration throughout lactation grew up until forty days postpartum. After this period, there was no cDNA amplification. In Nested PCR, positive results were detected in samples corresponding to the day of calving, 15th days, 20th days and 30th days postpartum, only.

Discussion: cDNA was detected in samples from one positive female, during forty days postpartum, but not on the 30th. On the other hand, amplified cDNA was observed on 30th day by nested-PCR. In this case, a false negative result was observed after real-time PCR, probably because sample corresponding to 30th days may not have been properly homogenized, so that the fraction used in real-time PCR was not representative. A higher number of positive samples were expected due to the higher sensitivity of the technique used. The low viral concentration in the milk due to high antibody titers, for example, leaded to a small number of cells containing the agent, reducing the possibility of detection. cDNA was not detected in any sample from three infected females. A possible false-positive serological reaction or the very low viral concentration in milk samples could explain the negative results, although some animals might be infected by a strain that could not be recognized by PCR.

Keywords: CAEV, small ruminant, milk, virology, Real-Time PCR.
INTRODUCTION

According to FAO (2014), there are 875 million of goats worldwide. Brazilian flock comprises approximately 14 million of animals, stating Brazil as the 18th in the exportation ranking [12].

Small ruminants can be infected by lentiviruses, such as Maedi-Visna Virus (MVV) and Caprine Arthritis-Encephalitis Virus (CAEV) [1]. Caprine arthritis encephalitis (CAE) is an infection that affects multiple systems such as locomotor, reproductive, respiratory and nervous, with clinical signs such as weight loss, arthritis, encephalitis and mastitis [14]. It is characterized by high prevalence of seropositive and generally low morbidity. The main route of transmission is via ingestion of contaminated colostrum and milk [1,5], although vertical transmission can occur [9]. The diagnosis of CAEV can be accomplished by serological and molecular reactions. The techniques widely used are Agarose Gel Immunodiffusion (AGID) and Enzyme-Linked Immunosorbent Assay (ELISA), recommended by the World Organization for Animal Health (OIE) and adopted by the Ministry of Agriculture (MAPA), in Brazil. Recently, several studies for molecular detection of CAEV in milk, using conventional PCR and real-time PCR (qPCR) are being carried out [7,10,15] because of their high sensitivity and specificity, besides the rapid results. Considering the elimination of CAEV through the milk of infected animals [7] and the importance of this virus in the goat production, the aim of this study was to evaluate the elimination pattern of CAEV in milk, assessing the frequency and the concentration eliminated during the lactation of experimentally infected goats.

MATERIALS AND METHODS

Animals, feeding and facilities

A cohort of four negative females for CAEV was inseminated with semen experimentally infected with CAEV-Cork strain. They were located in stalls at the Hospital of Ruminants from School of Veterinary Medicine and Animal Science from University of São Paulo, Brazil. Goats received coast-cross hay, pellet feeding, mineral salt and water ad libitum. All females were observed every day during pregnancy. After lambing, kids received warm bovine colostrum and bovine milk powder during two months.

Milk collection

Forty milk samples were collected in a five day interval during two months. Ten milliliters of milk of each teat were placed in Falcon tubes and frozen at -4°C until processing.

DNA extraction

Before cDNA extraction, a mixture of five milliliters from each teat (totaling 10 mL) was added in a Falcon tube. Samples were centrifuged at 1500 x g at 4°C for 10 min. Fat and supernatant were discarded and pellet resuspended in one milliliter of PBS solution. The suspension was transferred to another tube and nine milliliters of PBS was added. The suspension was centrifuged at 1500 g at 4°C for 10 min. The supernatant was discarded and the pellet resuspended in 200 µL of PBS. Samples were conditioned at -20°C.

cDNA extraction was performed using DNA Mini Kit (QIAGen) according Brinkhof et al. [2].

Real-Time PCR

Real-Time PCR (qPCR) was performed on Applied Biosystems 7500 Thermocycler [15]. All samples were analyzed in duplicate. Initially, the study of endogenous control for research of the constitutive gene (12S) for goats (called housekeeping), was performed using the following oligonucleotides - 12S-R 5’-TGAGTTTCGGGCTGTTGCCG-3’ and 12S-F 5’-CGAGCCACCGCGGTCATACG-3’. Real-time PCR reaction were prepared in 25 µL total volumes, containing 12.5 µL of master mix (Syber Green)1, 20 mM of each primer, 6.5 µL dH2O and 2 µL of sample DNA. Using positive samples for the first reaction, another reaction was performed using primers for lentiviruses - CF2 5’-GTTCCAGCAACTGCAAACTCAGTAGCAATG-3’ and CR2 5’-ACCTTTCTGCTTCTTCATTAAATTTCCC-3’, which were built based on the gag gene (conserved in retroviruses) [11]. The latter reaction was performed using the same conditions as described before.

In all reactions, a negative control consisting of the reagents excepting the DNA sample was used. The positive control was a CAEV cDNA (15 pmol/µL) kindly provided by Laboratory of Animal Virology from the Federal Rural University of Pernambuco. Both controls were used in duplicate. The standard curve was prepared using the cDNA diluted from 1:1 until 1:1000.

Nested-PCR

Nested PCR was performed in Thermal Cycler (Eppendorf) using positives samples detected by Real-time PCR. All reactions were prepared in 50 µL
total volumes, containing: 10X PCR buffer (200 mM Tris-HCl (pH 8.4), 500 mM KCl, 50 mM MgCl2, 10 µM of each dATP, dGTP, dTTP and dCTP, 10 µM of each primer, and 1U Taq Polymerase. After 5 min initial

denaturation at 95°C, 35 cycles were performed with
the following parameters: 95°C for 45 s, 55°C for 45 s
and 72°C for 45 s. The final extension was performed
at 72°C for 8 min. Fragments were analyzed by elec-
trophoresis in a 1.2% agarose gel in TBE buffer (0.09
M Tris-Borate and 0.002 M EDTA, pH 8.0), stained
with ethidium bromide. Bands were analyzed by UV
light and photographed using a photo documenter
(BioRad®)4.

RESULTS

After Real-Time PCR, cDNA was detected in
samples from one female, corresponding to the day of
calving, 14th, 20th, 25th, 35th and 40th day postpartum
(15% = 6/40). The absence of amplified cDNA in thirty
days postpartum, as well as in the final twenty days
of lactation, was observed. Sample corresponding to
the 7th day postpartum was not obtained. The virus
concentration throughout lactation grew up until forty
days postpartum. After this period, there was no cDNA
amplification (Figure 1).

In Nested PCR, positive results were detected
in samples corresponding to the day of calving, 15th
days, 20th days and 30th days postpartum, only.

![Figure 1. Evolution of cDNA (pmol) volume during lactation of healthy females inseminated with infected semen.](image)

DISCUSSION

C DNA was detected in samples from one
positive female, during forty days postpartum, but not
on the 30th. On the other hand, amplified cDNA was
observed on 30th day by nested-PCR. In this case, a
false negative result was observed after real-time PCR,
probably because sample corresponding to 30th days
may not have been properly homogenized, so that the
fraction used in real-time PCR was not representative.
Despite the absence of the sample corresponding to the
7th days postpartum, the trend of the results suggests
high probability that this sample was also positive.

In the present study, the number of positive
samples was higher compared to the results described by
Silva et al. [15], which detected positive samples by real-
time PCR in only 6% (11/179). However, the number of
positive samples in the present study was lower compared
to those described by Leroux et al. [10], which detected
100% (28/28) of positives by real-time PCR. Similarly,
De Regge et al. [4] detected 61% of positives by real-
time PCR after re-test positive samples on ELISA-test.
A higher number of positive samples were expected due to
the higher sensitivity of the technique used. The low viral
concentration in the milk due to a high antibody titers, for
example, leaded to a small number of cells containing the
agent, reducing the possibility of detection [14].

cDNA was not detected in any sample from
three infected females. Similarly, De Regge et al. [4]
did not detected amplification in 39% (09/23) by real-
time PCR using positive samples on ELISA-test. A
possible false-positive serological reaction or the very
low viral concentration in milk samples could explain
the negative results, although some animals might
be infected by a strain that could not be recognized
by PCR. Although the technique identifies animals
carrying the lentiviruses before seroconversion, it is
reported that PCR has become less sensitive when
compared to serological tests after viremia, and the
combination of them are recommended for diagnosis
and disease control [3].

In nested PCR, cDNA was obtained in samples
 corresponding to the day of calving, 15th, 20th and 30th
days postpartum (10% = 4/40). Similarly, Gregory et al.
[8], detected only 6% of positivity using nested-PCR.
On the other hand, Gregory et al. [7] detected 22.2%
of positive milk samples using the same technique.

Brinkhof et al. [2] reported that the detection of
cDNA in milk samples may indicate a viral infection in
the mammary gland, and it agrees with the transmission
by contaminated colostrum and milk hypothesis [13].
The infection of goats by ingesting infected colostrum
and milk has been described elsewhere [1,5,6,9,15].
Therefore, feeding kids with milk of infected females
should be avoided.

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Manufacturers
1Qiagen. Valencia, CA, USA.
2Life Technologies. Carlsbad, CA, USA.
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Funding. São Paulo Research Foundation (FAPESP) [Process nº 2012/19324-8].

Acknowledgements. We are grateful to Faculdade de Medicina Veterinária e Zootecnia (FMVZ) - Universidade de São Paulo (USP) for the structural support and all students for helping us during the experiment.

Ethical approval. The experiment was approved and performed under the guidelines of Ethics Committee for Animal Use of Faculdade de Medicina Veterinária e Zootecnia from Universidade de São Paulo.

Declaration of interest. The authors report no conflicts of interest. The authors alone are responsible for the content of the paper.

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